GUYANA 636.089 IG115Zq V.III No.2 1983

ANIMAL DISEASE REPORT

ANTILLES ZONE

APRIL - JUNE, 1983

VOL. III No. 2



INTER-AMERICAN INSTITUTE FOR COOPERATION ON AGRICULTURE

P.O. BOX 10-1089, GEORGETOWN, GUYANA,

IICA # 2.282 v. 3 no. 2

1983

Digitized by Google

CONTENTS

	Page
Bluetongue Studies in the Caribbean	1
Press Release	5
African Swine Fever Project	7
Rabies	9
Tetanus	10
Brucellosis	10
Tuberculosis	11
Haemoparasites	11
Leptospirosis	12
Endoparasites	13
Infectious Poultry Diseases	14
Mastitis	15
Metritis	15
Mange	15
Blackleg	16
Swine Erysipelas	16
Screwworms	16
Reproductive Herd Health	16
Import Inspections - Trinidad & Tobago	17
Animal Disease Reporting Personnel	18



BLUETONGUE STUDIES IN THE CARIBBEAN

Neutralising Antibody Studies at A.V.R.I., Pirbright

by

W.P. Taylor and I.D. Gumm (May, 1983)

As a general principle cattle, sheep and goats show a type-specific neutralising antibody response following a primary infection with bluetongue. If paired sera can be obtained it can be expected that the specificity of the neutralising antibody response will give a clear indication of the infecting type. However, it must sometimes be expected to find low levels of cross-neutralising antibody (antibody neutralising types other than those causing infection) even after the prime infection in life. This does not negate the value of the "neutralising antibody diagnostic test" as cross-neutralising levels will be low with respect to specific-neutralising levels. In our experience cattle tend to produce cross-neutralising antibodies more frequently than sheep or goats. Difficulty may arise - again in cattle - in that in some individuals diagnostic antibody levels may be only slightly higher than non-specific levels.

In cattle we have claimed to be able to diagnose a second infection either along with or sometime after the first (Herniman et al, 1983), based on the appearance of a high level of neutralising antibody to a second type. Our ability to do this was based on the availability of sera collected at monthly intervals from individual animals. In general though, the level, valency and duration of cross-neutralising antibody may be expected to rise the more frequently an animal is exposed to bluetongue and ultimately it will be impossible to use the neutralising antibody approach to diagnose the virus types present in an area.

As we had no detailed knowledge of the present study area and are unaware if bluetongue activity is seasonal or on-going throughout the year, we attempted

to work with sera collected from young animals (not older than 18 months) in the hope of selecting animals that had undergone only one or two infections and in which a broadened response had not occurred. In general, this approach seems to have succeeded and it is noticeable in the occasional two year old animal included to test our theory, that the response is in fact much broader. However, in viewing the results, it is apparent that very few sera give a mono-specific diagnostic response of the sort described above. On the other hand, very few sera are given antibody levels to a large number of types and we would suggest that most sera have been collected that have been infected with only a few virus types.

In attempting to interpret these results we have adopted a new approach - that of looking for antibody clusters, arguing that overall cross-neutralising will probably be low and random and that by looking at antibody titres at higher levels, some indication of the infecting type(s) may appear. For example, if we look at the results from Suriname it is apparent that 6 cattle sera of the 9 tested have antibodies at above a titre of 1:20 against BTV 6, that BTV 14 antibodies are found in 8 animals. No other virus types gives these clusters. When the threshold for examination is raised to 1:30 the BTV 6, 14 and 17 clusters become even more apparent. Of further significance, when the same method is applied to the (less cross reactive) sheep sera, the same three types give clusters and when goat sera are examined clusters appear for type 14 and 17. This reinforces the impression that the method of interpretation has an epidemiological basis as it might be expected that the same virus types would be found infecting all three host species.

Adopting this approach, we have tabulated what appears to be the infecting BTV types in each species in each area studied.

Digitized by Google

ANTIBODY CLUSTERS TO SEROTYPES OF BLUETONGUE VIRUS RECOGNISED IN CATTLE, SHEEP AND GOATS IN THE CARIBBEAN REGION AND COUNTRIES OF SOUTH AMERICA:

	makeen district and the second	No. Samples	
Country*	Species	Examined	Antibody cluster(s) to BTV
Jamaica	Cattle	1	Insufficient results
	Sheep	3	Insufficient results
	Goats	15	6, 14, 17
Antigua	Cattle	10	6, 12, 14, 17
	Sheep	6	6, 17
	Goats	10	6, 14, 17
Barbados	Cattle	11	1
	Sheep	24	1, 6
Grenada	Cattle	8	6, 14, 17
	Sheep	15	6, 17
	Goats	16	6, 14, 17
Trinidad	Cattle	56,	6, 14, 17
& Tobago	Sheep	17	14, 17
	Goats	20	6, 14, 17
Guyana	Cattle	9	14, 17
	Sheep	8	14
	Coats	6	Insufficient results
Suriname	Cattle	9	6, 14, 17
	Sheep	10	6, 14, 17
	Goats	10	14, 17

^{*}Countries listed by latitude from north to south

Number of sera tested

	Cattle	Sheep	Goats
¥		_	
Jamaica	1	3	15
St. Kitts/Nevis	1	~	4
Antigua	10	6	10
St. Lucia	No age:	s known not	tested
Barbados	11	24	
Grenada	8	15	16
Trinidad & Tobago	56	17	20
Guyana	9	8	6
Suriname	9	10	10
		ettip essin qilibi ugga danla	
	105	83	81

TOTAL: 269

CONCLUSION

In certain areas and in certain species there is good evidence in individual animals for recent infection with either BTV 6 or BTV 14; no similar evidence for BTV 17 exists but antibody to this type is ubiquitous. It is suggested that all three BTV types have been prevalent in the Caribbean during 1981 or the first half of 1982. It is impossible to determine the factors that control the spread of BTV in this area but it is quite striking that, with the possible exception of Barbados, the same viruses have been distributed throughout the entire study area in the course of a year. Further studies should be undertaken to determine the time of virus onset in each area.

REFERENCES

Herniman et al (1983) J. Hygiene.

PRESS RELEASE

Bluetongue Studies in the Caribbean

Bluetongue is a virus disease of ruminants transmitted by midges. In some areas of the world like the Caribbean, bluetongue virus is of little clinical concern. To date no clinical cases have been confirmed in the region. The potential of the disease, however, to cause epidemics, has led to the imposition of severe restrictions on the international movement of breeding livestock and germ plasm. Such restrictions led to the loss of exports of sheep from Barbados to Canada in 1972 and recently to Mexico. Jamaica has had a cancellation of a shipment of bovine semen to Nigeria. Caribbean countries wish to expand ruminant livestock development which focus on the Barbados Blackbelly Sheep, the developed cattle breeds of Jamaica and the Trinidad Water Buffalo. A policy governing the movement of ruminants within the Caribbean is required.

At the Regional Meeting of the Directors of Animal Health of the Antilles Zone held under the sponsorship of the Inter-American Institute for Cooperation on Agriculture in Barbados in 1981, it was decided to begin a study of the epidemiology of Bluetongue Virus in the Zone.

The first objective of the study, a serological survey to determine the prevalence of antibody to the common group antigen of Bluetongue Virus in cattle, sheep and goats from the region, was undertaken and executed by the Ministry of Agriculture, Food and Consumer Affairs, Barbados, at the Veterinary Diagnostic Laboratory, the Pine, St. Michael, in collaboration with consultants from the Centre of Tropical Animal Health, Department of Veterinary Medicine, University of Florida, USA.

The serological survey involving 500 sera from animals in participating countries showed overall prevalences of 70% in cattle, 66% in sheep and 76% in goats. Within countries, the prevalences were Jamaica 77%, St. Kitts/

Nevis 70%, Antigua 76%, St. Lucia 82%, Barbados 61%, Grenada 88%, Trinidad & Tobago 79%, Guyana 52% and Suriname 84%.

Following these results, 269 positive samples were tested at the Animal Virus Research Institute, Pirbright, England, in order to determine which of the 22 known serotypes were involved. Sera from young animals (not older than 18 months) were selected using an approach of looking at clusters of neutralising antibody titres at higher levels. Bluetongue Virus clusters 6, 14, 17 were disclosed throughout the region and in all three host species.

The conclusion was reached that in certain species there is good evidence in individual animals for recent infection with either Bluetongue Virus 6 or Bluetongue Virus 14. No similar evidence for Bluetongue Virus 17 exists but antibody to this type is ubiquitous. It is suggested that all three Bluetongue Virus types have been prevalent in the Caribbean during 1981 and early 1982. It is quite striking that the same viruses have been distributed throughout the entire study area in the course of a year. Further studies should be undertaken to determine the time of virus onset in each area. to identify the transmitting midges and to attempt virus isolation.

At a recent meeting in Kissimmee, Florida, USA, during May, 1983, a consultation was held with the leading authorities on Bluetongue Virus disease. There was unanimous opinion that the consistent results show that there would be little or no justification for restricting the movement of animals within the Caribbean Area, because of Bluetongue. They recommended that the respective Governments of the Region should be so advised and that research as indicated should be pursued.

In St. Vincent and Barbados, the serotypes disclosed by previous analysis were 1, 2, 4, 14, 15 and 1, 2, 5, 6, 7, 10 respectively. In the light of the results shown by the antibody cluster technique, sera can be retested accordingly.

The virus isolates present in the USA are 2, 10, 11, 13 and 17.

AFRICAN SWINE FEVER PROJECT

			Compensation Paid
Country	Size	No. Slaughtered	May, 1982 - June, 1983
Haiti	Adults	168007	6,720,280
¥.	Young	116444	2,328,880
	Piglets	99940	499,700
	TOTAL	384391	9,548,860
		man of the company of	- Contract of the Contract of

Laboratory Report - April - June, 1983

Week	No. of Blood Samples	No. Positive	% Positive
April 4-8	778	34	4.4
9-15	772	42	5.4
16-22	633	12	1.9
23-30	82 5	15	1.8
May 1-6	826	24	2.9
7-13	429	11	2.6
14-20	733	27	3.7
21-27	349	2	0.6
May 28-June 3	739	23	3.1
June 4-10	860	45	5.2
11-17	353	9	2.6
18-24	1120	41	3.7
25-31	1591	7.5	4.7
	10008	360	% 3.6

Raking Operations

					No. of
<u>Date</u>	Chefs de Section	Pigs Captured	Claimed	Unclaimed	Owners
June 1983	249	233	199	34	95

Cleaning and Disinfection

	April - June 1983	Sept 1982 - June 1983			
Slaughter/Compensation Sites	147	438			
Piggeries	2	7			
Abbatoirs	16	42			
Positive Cases Sites	75	224			
Markets	31	92			
	271	803			
	<u> </u>				

Epidemiological Surveillance

	April - June 1983	August 1982 - June 1983
Roads		
Vehicles Inspected	26557	77388
Vehicles Disinfected	1295	3134
Meat Seized (kg)	216825	6236.25
Maritime		
Boats Inspected	1843	6258
Boats Disinfected	24	230
Meat Seized (kg)	494.5	677.5 + 2pigs
Airport		
Aircraft Inspected	129	252)
Meat Seized (kg)	23	252) 89.5) Feb - June 1983

	April - June 1983	August 1982 - June 1983
Haitian/Dominican Border		
Vehicles Inspected	1071	1357 19) Feb - June 1983
Vehicles Disinfected	19	19 Feb - June 1983
Meat Seized (kg)	627.25	1256.5 + 2 Pigs

Turks & Caicos Islands

All fourteen sera (14) submitted were negative for African Swine Fever by the Indirect Fluorescent Antibody Test. Ten (10) sera were positive for Hog Cholera by the IFA Test and the FA Neutralisation Test. Positive Hog Cholera sera were obtained from all islands surveyed.

(USDA Report - Following Visit - Eskew/Alexander, May 1983)

RABIES

		No. of Cases	Cumulative	No. Vaccinated	Cumulat	ive
Country	Species	/Quarter	Total	/Quarter	<u>Total</u>	
Grenada	Bovine Canine	-	3	13 45	35 181	
(April Report Only)	Caprine Ovine		6	14)	83	299
Suriname	Bovine	5	5	-	-	5
Trinidad & Tobago	Bovine Buffalo Caprine Equine		-	1026 19 433 6	1781 19 586 25	2450
	Ovine			22	47	2458

TETANUS

Country	Bov	ine	Capr	ine	Donl	кеу	Equ	ine	Ovi	<u>ie</u>	Porc	ine	Tot	<u>a1</u>
	Q	С	Q	С	Q	С	Q	С	Q	С	Q	С	Q	С
Grenada	-	-	4*	12*	-	-	1	2	4*	12*	2	8	7	22
Guyana	_		-	-	3	3	8	12	5	18	-		16	33
Jamaica	3	6	5	35	-	-	10	14	-	-	26	40	44	95
Trinidad & Tobago	1	1	11	15	2	2	-	1	-	-	-	-	14	19

* Grouped Data

BRUCELLOSIS

		No. Tested/Quarter		No.	No.	Cumu	lative	
Country	Species	Slaughter	Field	Farms	Pos	Slaughter	<u>Field</u>	Pos
Barbados	Bovine	-	3	_	-	_	6	_
Jamaica	Bovine	8483	5402	85	4	18907	12724	22
St. Lucia	Bovine	-	-	-	-	-	50	-
Suriname	Bovine	-	335	5	3	91	610	7
Trinidad	Bovine	-	2	-		_	2	-
& Tobago	Ovine		No.	-	-	-	12	-

TUBERCULOSIS

		No. Tested	No.	No.	Cumulati	.ve
Country	Species	/Quarter	Farms	Positive	No. Tested	No. Pos.
Jamaica	Bovine	3916	46	0	9085	4
St. Lucia	Bovine		-	<u></u>	50	-
Suriname	Bovine		••		1	1
Trinidad	Bovine			-	25	-
& Tobago						

HAEMOPARASITES

		No. (Cases	No.	No.	No.	Cumula	tive No.
Country	Species	/Quai	ter	Deaths	Tested	Farms	of Cas	es
		Ana	Piro				Ana	Piro
Barbados	Bovine	ima	-	-	***		33	4
	Canine	***	_	•••		-	***	1
	Caprine	_	-	-44	_	-	15	_
	Equine	_	~	-	~		-	3
	Ovine		-		_	~	1	-
Grenada	Bovine	2	-	-	2	2	. 9	-
Guyana	Bovine	17	8	7	-	-	118	36
	Ovine	8	•••	-	-		29	-
Jamaica	Bovine	21	***			21	46	-
	Canine	_	7	eos:	7	7	_	20
	Caprine		-	ame.	-	-	1	
Trinidad	Bovine	251	4	9	-	-	439	11
& Tobago	Caprine	1	-	ades.			-	-

LEPTOSPIROSIS

		No. Tested/	No.	Cumu	lative
Country	Species	Quarter	Positive	No. Tested	No. Positive
Barbados	Bovine	12	12	13	12
	Canine	9	8	11	8
	Equine	12	12	13	13
O	Danida a			504	220
Guyana	Bovine	-		594	339
	Human	29	11	78	22
Jamaica	Bovine	1132	588	1593	742
	Canine	54	19	89	31
	Caprine	20	4	116	40
	Human	120	57	224	83
	Ovine	3	0	13	1
	Porcine			1	-
Trinidad	Bovine	2	_	2	_
& Tobago	Ovine	12	· 	12	-

ENDOPARAS I TES

Bovine Q Canine Q Caprine Q Equine Q Feline Q Ovine Q Porcine Q Zoo Q Total Q (21)11 17 (86)55 117 (19)17 22 (8)8 12 (4)2 3 (17)15 39 (1)1 1 - 109 211 9 240 39* 112* 249* 1011* - 20 39* 112* 249*1011* 79 357 - 457 1821 19 1659 35 75 - 372 8 20 5 5 5 322 3 522 5 14 80 3036 617 1469 596 1197 53 131 296 613 462 981 2024 4391
Feline Ovine Porcine Zoo 1 Q C Q S 3 S 22 S 14 S S S 3 S 22 S 14 S
Feline Ovine Porcine Zoo 1 Q C Q S 3 S 22 S 14 8 S S 3 3 15 - - 2 20 -
Feline Ovine Porcine Zoo 1 Q C Q A D A D A D A D A D A D A D A D A D D
Feline Ovine Porcine Zoo 1 Q C Q A 2 A S S S S S S S S S S S S S S S S
line Ovine Porcine Zoo 1 C Q C Q C Q C (3 (17)15 39 (1)1 1 103 112* 249*1011* 79 357 45 5 5 322 3 522 5 14 8 - 296 613 462 981 20 - 8 39 3 15 3
Ovine Porcine Zoo 1 Q C Q C Q C (17)15 39 (1)1
orcine Zoo 1 C Q C (1 109 357 45 522 5 14 8 981 20
orcine Zoo 1 C Q C (1 109 357 459 522 5 14 8 981 20
orcine Zoo 1 C Q C (1 109 357 45 522 5 14 8 981 20
Zoo 1 Q C (109 45 20 20
L C C 211 321 321 391

() Total Tested

Grouped Data e.g., Dogs & Cats

Sheep & Goats

Includes Buffaloes

,	No. of Birds Affected	No. of Farms Affected	Trinidad & Tobago	Suriname No. of Birds Affected	No. Vaccinated	No. of Birds Affected	No. of Farms Affected	Jamaica	Mortality	No. of Birds Affected	No. of Farms Affected	Guyana	INFECTIOUS POULTRY DISEASES Country As CRD
	56000	11		ı	ı	ł	ı		í	1	ŧ		Aspergillosis Dive
	273800	82		1	1	1 37	ŧ		49	49	ı		CRD &S
	273800 1480 120	ω		l	1	37500	69		37	78	ŧ		Coryza
	1	j4		ı		i	ł		1	1	ì		Encephalomalacia
* In o Re	61300	14		10	1	14500	12		50*	6903	1		Endoparasites
Includes Reported	ı	1		į		t	i		ı	ı	1		Fowl Pox
23 Syn	200	 -		ı	1	ı	ı		í	1	ı		Gumboro Disease
₽ħ	1	ı		I	61000	ı	47		f	ı	ì		Infectious Bronchitis
	ı	ı		t		1	I		í	ı	ı		Leucosis
Histomoniasis hea in import	10400	11		ı	200000	t	ł		7	7	1		Marek's Disease
niasis imported	1	ı		t	ı	t	1		1	1	1		Mycoplasmosis
guinea Fowl	49700	ω		1	6700000	i	1	A THE PERSON NAMED IN THE	ı	ı	1		Newcastle Disease
Fowl	1	ı		1	1	ī	ı		1	1	ı		Pasteurellosis
,	10000	-		t	1	ı	t		1	ı	1		Salmonellosis
				e .									I

					MANGE			METRITIS				MASTITIS	;	
	Trinidad & Tobago	Jama ica	Guyana	Grenada	Barbados	Trinidad & Tobago	Jamaica	Guyana	Trinidad & Tobago	Jamaica	Guyana	Grenada		Country
	i	22	1	ı	į	82	289	ı	184	484	246	15	Q	Во
	9	39	1	ı	1	210	465	ı	383	882	266	37	C	Bovine
	ı	1	ı	4	1	ŧ	į	1	1	ı	ı	ı	Q	Can
	ı	i	9	13	4	ŧ	i	1	ı	í	í	ì	C	Canine
* 0	ŧ	ŀ	∞	i	í	iл	86	1	14	24	11	13*	Q	Cap
Grouped Data	r	1	∞	i	ŀ	9	120	t	25	47	20	49	C	Caprine
Data	í	2		ŧ	ŧ	1	i	ŧ	1	1	í	i	Q	Equine
	ı	2		l	ı	1	í	1	ŧ	1	. 1	1	C	ine
	ŧ	ì	i	ı	ł	1		ı	i	i	ı	ı	Q	Feline
	1	ı	ł	i	1	ŧ	1	ŧ	į	l	ı	ı	C	ne
		1	i	2	1	 -	21	ı	ı	i	1	13*	0	Ovine
		ı	i	2	1	-	25	ŧ	ŧ	ì	ı	49	C	ne
		25	23	ı	ı	19	40	1	ı	18	72		0	Porc
	28	48	50	Ĺ	ı	ယ္	63	į	j4	36	72	27	C	Porcine
	2	49	32	6	i	107	436	43	198	526	329	39	Q	To
	3 8	89	59	18	4	253	674	i	409	965	358	113	C	Total

BLACKLEG

Country	Species	No. of Cases	No. Vac	ccinated	No. of Farms		
			Q	C	Q	С	
Jamaica	Bovine	-	8170	12497	1706	2137	

SWINE ERYSIPELAS

Country	No. of	Cases	No. Va	No. of Farms		
	Q	С	Q	С	Q	С
Jamaica	-	-	1732	2935	73	142
Trinidad & Tobago	37	60	-	-	-	-

SCREWWORMS

Country	Bovine		Caprine		Equine		<u>0v</u>	ine	Por	cine	To	Total	
	Q	С	Q	С	Q	C	Q	С	Q	С	Q	С	
Guyana	320	940	43	43	27	27	53	63	-	_	443	1073	
Trinidad & Tobago	9	21	2	5	_	2	-	1	4	5	15	34	

REPRODUCTIVE HERD HEALTH

Country	No. of Cows Examined		No. of Farms		No. Vac	_	No. c	of Farms	No . Exa	Bulls
	Q	C	Q	С	Q	С	Q	C	Q	С
Jamaica	4402	9101	103	194	3065	83	83	83	-	194
Trinidad & Tobago	755	2233	75	158	-		•••	-	-	-

IMPORT INSPECTIONS - TRINIDAD & TOBAGO

Coun	trv	Ωf
OUGH		~_

Origin	Avian	Bovine	Canine	Caprine	Equine	Ovine	Porcine	Turtles	Total
Barbados	هنت		25		22	-		-	47
Canada	-	-	1	***	3	-	18		22
England	200	494	50	-	3	-	_	-	253
Germany	-	-	1	-	-	-	_	-	1
Grenada				266		650	-	_	916
Guyana	106	-	-	-	-	-	-	-	106
Martinique	· -	-			7	-	_	2	9
Netherlands	3898		- was	-	-	-	-	-	3898
Peru	-		1	-	-	-	-	-	1
Puerto Rico	23	_	_		1	-	_	-	24
St. Lucia	3		-	-	-	-	-	-	3
St. Vincent			1	19	~	174	18	-	212
USA	44	97	9	44.	8	-	13	2070	2241
TOTAL	4274	97	88	285	44	824	49	2072	7733

ANIMAL DISEASE REPORTING PERSONNEL

Barbados: Dr. Trevor King

Senior Veterinary Officer Ministry of Agriculture, Food

& Consumer Affairs Animal Health Services The Pine, St. Michael

Dr. Vincent St. John Veterinary Pathologist Veterinary Laboratory

St. Michael

Dr. W.M. Christian Dominica:

> Chief Veterinary Officer Ministry of Agriculture Roseau, Commonwealth of

Dominica

Grenada: Dr. K.S. Manyam

> Chief Veterinary Officer Ministry of Agriculture, Rural

Development & Cooperatives P.O. Box 141, St. George's

Tanteen

Guyana: Dr. R.N.D. Raja

Principal Agriculture Officer (Ag.)

Ministry of Agriculture

Veterinary & Livestock Division

Regent & New Garden Sts. G'town

Haiti: Dr. Fred Calixte

Chef, Service Veterinaire

DARNDR

Damien, Port-au-Prince

Jamaica: Dr. Clifford L. Grey

Director of Veterinary Services (Ag.) Deputy Director

Ministry of Agriculture P.O. Box 309, Kingston

St. Lucia: Dr. Keith Scotland

Chief Veterinary Officer Ministry of Agriculture

Castries

Dr. Robert Lieuw-a-Joe Suriname:

> Chief, Veterinary Inspection Ministry of Agriculture

P.O. Box 1016, Paramaribo

Trinidad Dr. Vincent G. Moe

Technical Officer - Livestock Div. & Tobago:

Ministry of Agriculture

St. Clair Circle, Port of Spain

Miss Judy Baldeau Laboratory Technician

Ministry of Agriculture, Rural Development & Cooperatives P.O. Box 141, St. George's

Tanteen

Dr. Lennox Applewhaite Senior Veterinary Officer Veterinary Laboratory Ministry of Agriculture

Mon Repos, East Coast Demerara

Dr. Max Millien

Chef, Laboratoire Veterinaire

DARNDR

Damien, Port-au-Prince

Dr. George Hylton

Linton McDonnough Memorial Veterinary Laboratory P.O. Box 309, Kingston

Mr. John Simon

Laboratory Technician Ministry of Agriculture

Castries

Dr. Ken Moll

Veterinary Diagnostic Laboratory

P.O. Box 1016, Paramaribo

Dr. Edward P. Cazabon Veterinary Pathologist

Veterinary Diagnostic Laboratory

Curepe, Port of Spain